

# The endothelium in acute lung injury/acute respiratory distress syndrome

Nikolaos A. Maniatis<sup>a</sup> and Stylianos E. Orfanos<sup>a,b</sup>

<sup>a</sup>M. Simou<sup>1</sup> Laboratory, University of Athens Medical School, Athens, Greece and <sup>b</sup>2nd Department of Critical Care, Attikon Hospital, University of Athens Medical School, Athens, Greece

Correspondence to Dr Stylianos Orfanos, 2nd Department of Critical Care, Attikon Hospital, 1, Rimini St., 124 62 Haidari, Athens, Greece  
Tel: +30 210 723 5521; fax: +30 210 723 9127;  
e-mail: sorfanos@med.uoa.gr

**Current Opinion in Critical Care** 2008, 14:22–30

## Purpose of review

Since pulmonary edema from increased endothelial permeability is the hallmark of acute lung injury, a frequently encountered entity in critical care medicine, the study of endothelial responses in this setting is crucial to the development of effective endothelial-targeted treatments.

## Recent findings

From the enormous amount of research in the field of endothelial pathophysiology, we have focused on work delineating endothelial alterations elicited by noxious stimuli implicated in acute lung injury. The bulk of the material covered deals with molecular and cellular aspects of the pathogenesis, reflecting current trends in the published literature. We initially discuss pathways of endothelial dysfunction in acute lung injury and then cover the mechanisms of endothelial protection. Several experimental treatments in animal models are presented, which aid in the understanding of the disease pathogenesis and provide evidence for potentially useful therapies.

## Summary

Mechanistic studies have delivered several interventions, which are effective in preventing and treating experimental acute lung injury and have thus provided objectives for translational studies. Some of these modalities may evolve into clinically useful tools in the treatment of this devastating illness.

## Keywords

ALI, ARDS, endothelium, lung

Curr Opin Crit Care 14:22–30  
© 2008 Wolters Kluwer Health | Lippincott Williams & Wilkins  
1070-5295

---

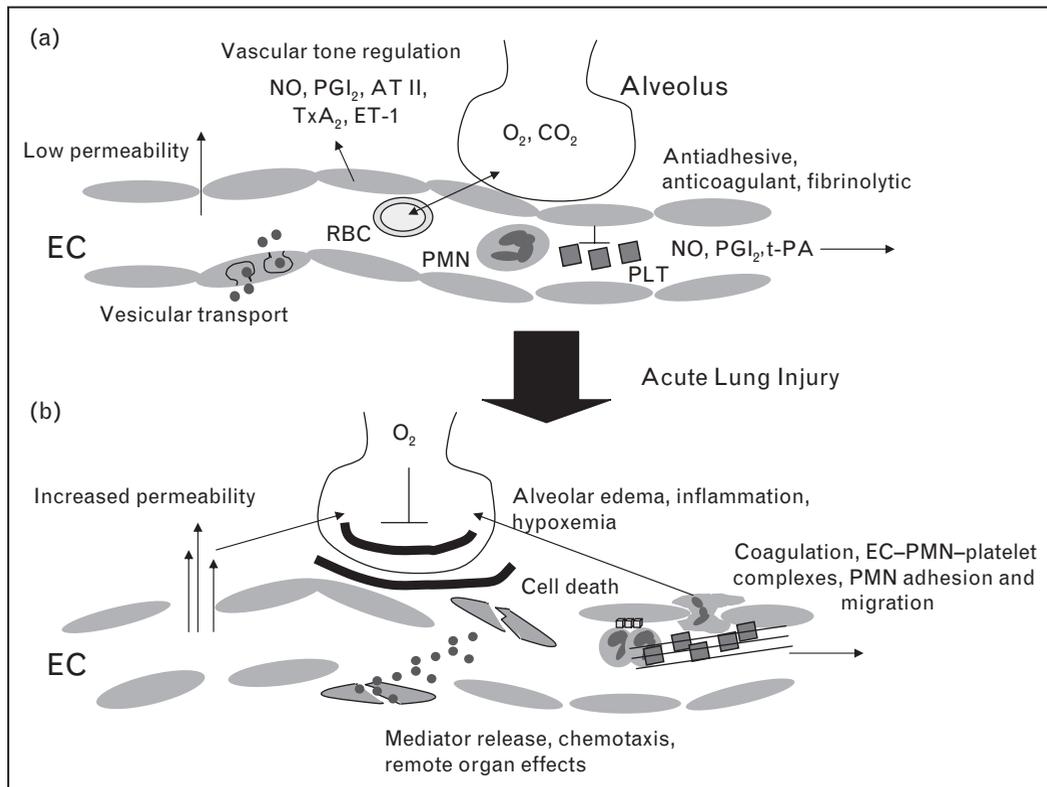
## Introduction

The intimal lining of all blood vessels is a single layer of functionally and structurally heterogeneous endothelial cells depending on organ and vascular bed location [1]. Pulmonary microvascular endothelium is a metabolically active organ essential for maintaining adequate pulmonary and systemic cardiovascular homeostasis [1,2]. Noxious stimuli compromise pulmonary endothelial functional and structural integrity leading to noncardiogenic pulmonary edema and parenchymal inflammation. Thus, pulmonary endothelium has a key role in the development of acute lung injury (ALI) and its most severe form, the acute respiratory distress syndrome (ARDS). While no therapeutic breakthroughs in the lung endothelial field were reported over the past 12 months, several experimental contributions to the understanding of endothelial mechanisms of injury and repair were made. We will therefore focus on these recent advances and present some novel endothelial-based treatments.

---

## Major endothelial properties

Endothelial cells in lung vessels are arranged in a continuous monolayer forming a tight barrier [1]. Major determinants of endothelial fluid permeability are intercellular protein bridges called tight and adherens junctions (AJ) and contact sites to the substratum called focal adhesions [3]. These structures allow paracellular exchange of very small amounts of water and solutes. An exquisitely developed system of vesicular carriers allows uptake of macromolecules into and transport across endothelial cells in a highly selective manner. Besides permeability regulation, pulmonary endothelial cells modulate vascular tone, ventilation–perfusion matching, hemofluidity, and interactions with blood-borne cells. Inflammatory states such as sepsis induce a phenotypic shift in endothelial cells with secretion of inflammatory and chemotactic substances, expression of adhesion molecules, increased permeability and loss of anticoagulant functions [2]. A schematic representation of major lung endothelial functions in health and

**Figure 1 Schematic illustration of major pulmonary vascular endothelial functions in health (a) and in acute lung injury (b)**

(a) Pulmonary vascular endothelium forms a single-layer interface separating blood from the extravascular space. The layer is composed of tightly connected endothelial cells (ECs) and allows minimal filtration of fluid, thus preventing alveolar flooding and facilitating gas diffusion. An extensive network of vesicles is responsible for the highly selective, receptor-mediated uptake and transport of macromolecules (e.g. albumin) across the barrier and the secretion of macromolecules (e.g. von-Willebrand factor) in the blood. Lung endothelial cells are largely responsible for regulating vascular smooth muscle (VSM) tone, maintaining low pulmonary vascular resistance and appropriate ventilation-perfusion matching. This is achieved by balancing the secretion of vasodilators also possessing antiproliferative and anti-inflammatory effects (e.g. nitric oxide, NO; prostacyclin, PGI<sub>2</sub>) against that of vasoconstrictors also inducing cell proliferation/hypertrophy and inflammation (e.g. thromboxane A<sub>2</sub>, TxA<sub>2</sub>; angiotensin II, AT II; endothelin-1, ET-1). Intravascular clotting is prevented by agents counteracting platelet adhesion such as nitric oxide and prostacyclin and lysis of endovascular thrombi is facilitated by fibrinolytic agents such as tissue plasminogen activator (t-PA). (b) The exposure of lung endothelial cells to noxious stimuli, including bacterial toxins, inflammatory mediators or excessive mechanical forces triggers a shift towards a 'pro-inflammatory' and thrombophilic endothelial cell phenotype, characterized by platelet and leukocyte adhesion molecule expression, leading to tissue infiltration and intravascular clotting. These processes are augmented by endothelial cell apoptosis and necrosis, which produce further inflammatory mediators and allow clot formation on denuded surfaces. Endothelial cell cytoskeleton contraction, triggered by inflammatory mediators (e.g. thrombin), increases endothelial permeability. The ensuing plasma extravasation leads to pulmonary edema, the hallmark of acute lung injury and the acute respiratory distress syndrome. ; PLT, platelet; PMN, polymorphonuclear leukocyte; RBC, red blood cell.

disease is given in Fig. 1. For a more detailed analysis of pulmonary endothelial metabolic properties and endothelial cell contribution to ALI/ARDS pathophysiology, the reader is referred to [2].

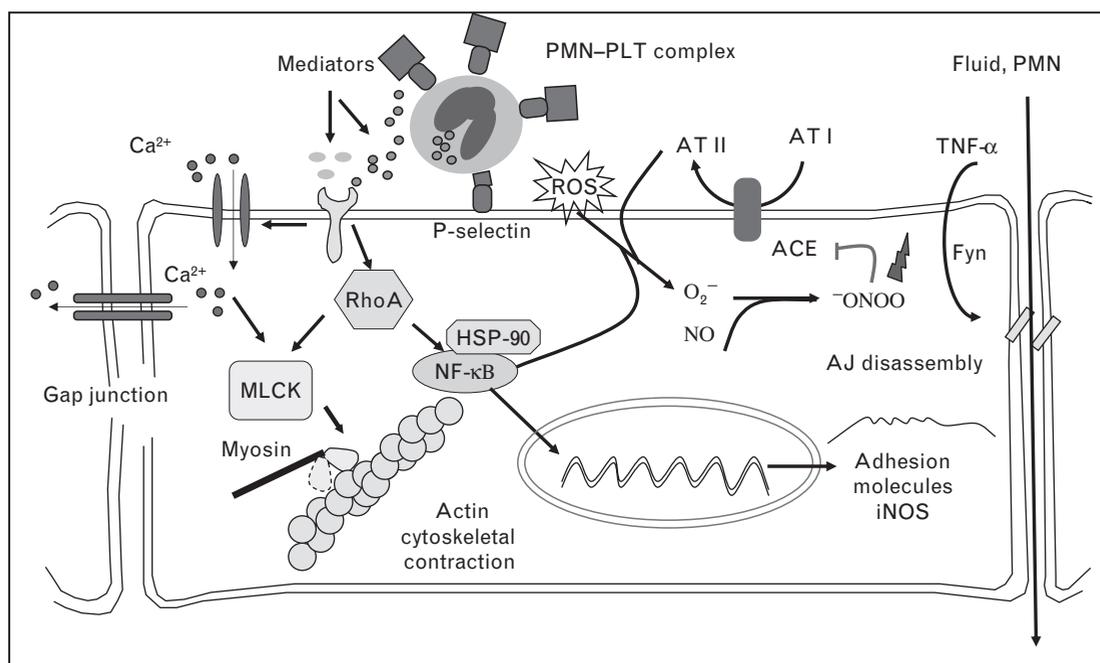
### Mechanisms of endothelial barrier disruption in acute lung injury

Endothelial barrier disruption is a *sine qua non* of ALI/ARDS pathogenesis; related pathological mechanisms are presented in Fig. 2, along with additional proinflammatory endothelial cell pathways implicated in lung injury.

#### Cytoskeleton

Endothelial cell actin cytoskeleton is a complex network of actin molecules polymerized into filaments. Multiple

connections of the cytoskeleton to junctional proteins, the glycocalyx, and focal adhesions exist. Stimulation of endothelial cells by agonists, such as thrombin, signals actin filament rearrangement into thicker actin cables or stress fibers, and enhances actin-myosin interactions. Cytoskeletal contraction, facilitated by the motion of myosin along actin fibers, generates tension that pulls cells away from each other creating inter-endothelial gaps and increasing permeability. Myosin light chain (MLC) phosphorylation by MLC kinase (MLCK) augments cytoskeletal force generation [4]. Small GTPase-induced activation of MLCK and other cytoskeletal-remodeling compounds, such as p21-activated kinase, lead to pulmonary endothelial barrier compromise and assist leukocyte migration in sepsis [5]. Genetic deletion or pharmacologic inhibition of MLCK210, the endothelial

**Figure 2 Schematic illustration focusing on recently described endothelial-based molecular aspects of acute lung injury (ALI) pathogenesis**

Binding of inflammatory mediators, including thrombin, thromboxane  $A_2$  and transforming growth factor- $\beta$  on their receptors on endothelial cell (EC) surface activates a cascade of signaling events that lead to endothelial barrier disruption and modulation of gene expression. Sources of these mediators include platelets (PLTs), leukocytes and other tissue cells. Calcium ion ( $Ca^{2+}$ ) influx following endothelial cell surface receptor ligation raises  $Ca^{2+}$  levels, activating downstream targets and propagating the signal to adjacent endothelial cells via gap junctions. The small GTPase RhoA and its downstream effectors mediate actin stress fiber formation and contraction of the actin cytoskeleton, allowing leukocyte transmigration and plasma leakage in the extravascular space. Nuclear factor  $\kappa B$  (NF- $\kappa B$ ) activated by RhoA stimulates expression of leukocyte and platelet adhesion molecules. Molecular chaperone heat shock protein-90 (HSP-90) promotes NF- $\kappa B$  activation. Adherent leukocytes, primarily polymorphonuclear leukocytes (PMNs), release superoxide radicals ( $O_2^-$ ) with potent bactericidal and cytotoxic effects. Reactive oxygen species (ROS) bind with nitric oxide (NO) to form further aggressive molecules such as peroxynitrite ( $-ONOO$ ). Angiotensin converting enzyme (ACE) is critical in ALI pathogenesis by producing angiotensin II (AT II), which has proinflammatory actions. In addition to promoting NF- $\kappa B$  activity and apoptosis, tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) can directly disrupt endothelial cell adherens junctions (AJ) by activating tyrosine kinase Fyn, which subsequently phosphorylates various adherens junction protein components. MLCK, myosin light chain kinase; iNOS, inducible nitric oxide synthase;  $\rightarrow$ , activation;  $\perp$ , inhibition.

cell-specific MLCK isoform, protects mice from experimental pulmonary edema, inflammation and death [6]. Furthermore, activation of nuclear factor  $\kappa B$  (NF- $\kappa B$ ; i.e. a central transcription factor of inflammation-related genes) and expression of the inducible isoform of nitric oxide synthase (iNOS) were attenuated in endotoxemic MLCK210 knockout mice, translating into a reduction of nitric oxide, reactive oxygen species and vascular hyporeactivity, and pointing to a link between cytoskeletal and paracrine endothelial cell pathways [7 $\bullet$ ].

A second, less well studied component of the endothelial cell cytoskeleton, the microtubules, comprises a system of polymerized tubulin molecules in rod-shaped arrangements that extend from the center to the periphery of the cell. Microtubule destabilization by the experimental chemotherapeutic agent 2-methoxyestradiol increases endothelial cell permeability [8]. Conversely, inhibition of microtubule destabilization with paclitaxel leads to microvascular barrier enhancement in mice challenged with lipopolysaccharide (LPS) [9].

### Intracellular signaling

Inflammatory mediators bind to their endothelial cell receptors to activate signaling events culminating in cytoskeletal contraction and increased microvascular permeability. These ligands include thrombin, vascular endothelial growth factor (VEGF), transforming growth factor- $\beta$  and thromboxane  $A_2$  ( $TxA_2$ ). In many cases, a complex interplay between cell junctional elements and focal adhesions results in rapidly occurring actin cytoskeletal remodeling [10]. More delayed effects comprise the induction of enzymes favoring actin stress fiber formation such as small GTPase RhoA [11].

Injured tissues may serve as the origin of inflammatory mediators, promoting the expansion of inflammatory processes. The proinflammatory cytokine tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) is secreted by lung endothelial cells following ischemia/reperfusion, and induces pulmonary edema in the nonischemic contralateral lung [12 $\bullet$ ]. TNF- $\alpha$  may promote disruption of endothelial cell junctions by phosphorylation of AJ

proteins such as vascular endothelial cadherin and  $\beta$ -catenin [13<sup>•</sup>].

Intracellular  $\text{Ca}^{2+}$  is an important second messenger in pathways that signal permeability increases. Cytosolic  $\text{Ca}^{2+}$  concentration is the composite of calcium ions from intracellular stores and the extracellular space. A route of  $\text{Ca}^{2+}$  entry into endothelial cells via the vanilloid transient receptor potential channel was recently uncovered [14<sup>•</sup>]. Activation of this mode of  $\text{Ca}^{2+}$  influx increased permeability in isolated mouse lungs and produced disruption of the alveolocapillary membrane at the electron microscopy level. In addition to activating MLCK,  $\text{Ca}^{2+}$  can trigger the expression of cell surface adhesion molecules, enabling docking of circulating polymorphonuclear leukocytes (PMNs) to endothelial cells and their transmigration in the interstitium and airspace.  $\text{Ca}^{2+}$  signals can be propagated along interconnected endothelial cells via gap junctions [15<sup>••</sup>], explaining how inflammation can rapidly spread across large areas of the lung and become manifest as extended patches of consolidation in imaging studies.

NF- $\kappa$ B is an important transcription factor of genes induced in inflammatory states, including sepsis and ventilator-induced lung injury (VILI). Heat-shock protein 90 (HSP-90), an abundantly expressed molecular chaperone, is required for NF- $\kappa$ B uncoupling from its associated inhibitory I $\kappa$ B and consequent activation. By administering the HSP-90 inhibitors radicicol or a geldanamycin analog to endotoxemic mice, Chatterjee *et al.* [16<sup>••</sup>] reduced NF- $\kappa$ B activation, resulting in decreased lung inflammation and endothelial permeability, as well as in improved lung function, systemic homeostasis and survival.

### Endothelial interactions with blood-borne cells

An important endothelial cell proinflammatory function is the secretion of the potent chemotactic cytokine interleukin (IL)-8 [17]. This and other substances secreted by endothelial cells are implicated in causing cytoskeletal PMN alterations, thus reducing their deformability and promoting sequestration at sites of inflammation [18,19]. In addition, activated PMNs have been shown to directly induce vascular leak by free radical production [20<sup>•</sup>].

Platelet and PMN binding to endothelial cells is required for intravascular clotting and PMN transendothelial migration, respectively. Platelet P-selectin facilitates platelet-endothelial cell binding [21]. Bound platelets complexed with PMNs release mediators such as  $\text{TxA}_2$  and soluble CD40 ligand, leading to endothelial cell and PMN activation, respectively [22<sup>••</sup>,23<sup>•</sup>]. In a similar respect, PMN diapedesis across endothelial cells was found to be dependent on signaling pathways involving

endothelial cell G proteins, specifically  $\text{G}\alpha(i)$ , which transduce signals downstream of G-protein-coupled receptors [24<sup>•</sup>]. The interaction of adherent PMNs with endothelial cell AJ was studied by Orrington-Myers *et al.* [25<sup>•</sup>]. AJ disassembly by expression of dysfunctional vascular endothelial cadherin, the extracellular AJ component, reduced PMN migration, NF- $\kappa$ B activity and intracellular adhesion molecule (ICAM)-1 expression in LPS-treated mice. It seems, therefore, that the presence of intact endothelial cell AJ is required for proper PMN binding and migration.

### Reactive oxygen and nitrogen species

Oxygen and nitrogen-based free radicals including superoxide anion and nitric oxide (NO) are important biological endothelial cell signaling molecules but can cause tissue damage at high concentrations. Interventions depleting sources of superoxide anion such as xanthine oxidoreductase or aldehyde 4-hydroxy-2-nonenal reduce free radical production and the associated microvascular leak in various in-vivo and in-vitro injury models [26–28]. PMN-derived free radicals induce pulmonary endothelial cell dysfunction in the setting of transfusion-related acute lung injury: when blood containing antibodies to human neutrophil antigen 3a, expressed on the surface of recipient PMNs, is transfused to susceptible patients, PMN stimulation may elicit toxic effects on endothelial cells via reactive oxygen species (ROS) production [29<sup>•</sup>].

Catalytically active iron can induce oxidative stress in lung tissue. Such alterations in the redox state of lung endothelial cells [30] are accompanied by NF- $\kappa$ B activation leading to ICAM-1 expression, and downregulation of vascular endothelial cadherin expression.

Inhaled nitric oxide has been used in the treatment of ALI/ARDS to improve gas exchange and lower pulmonary vascular resistance, despite concerns for potential local or distant harmful effects [31<sup>•</sup>]. In a rat model of *Pseudomonas* pneumonia endothelial and epithelial permeability were separately quantified *in vivo* using a double flux technique of radio-labeled albumin. Inhaled nitric oxide was found to increase endothelial permeability [32] independent of tissue PMN infiltration, suggestive of a direct effect on pulmonary endothelium. This notion was corroborated in additional in-vitro and in-vivo studies, in which iNOS inhibition was associated with endothelial cell barrier protective effects [33,34].

### Renin-angiotensin-aldosterone system

The angiotensin converting enzyme (ACE)-I, responsible for the conversion of angiotensin I to angiotensin II is expressed primarily on the surface of pulmonary microvascular endothelial cells [1<sup>•</sup>]. Even though pulmonary

endothelial ACE activity reduction has been documented in various animal models of lung injury as well as in patients with ALI/ARDS [2], evidence is accumulating that ACE is directly involved in the pathogenesis of the disorder. It has been proposed that the above-mentioned ACE activity reduction may be related to enzyme downregulation mediated by overproduction of peroxynitrite and other ROS/reactive nitrogen species [2], as schematically presented in Fig. 2. ACE inhibition in rats appears to ameliorate oleic acid-induced ALI and VILI [35,36<sup>\*</sup>]. In the latter study, increased angiotensin II levels were detected in lung tissue following VILI, suggesting that ACE may contribute to lung injury via angiotensin II production. The same group followed Chinese ICU patients with ARDS and compared clinical outcomes among carriers of the three ACE genotypes: II, ID, DD. ARDS patients with the II genotype appeared to have a survival advantage over patients with the ID or DD genotype [37<sup>\*</sup>]. These findings are in agreement with previous reports showing that the D allele is associated with higher ACE activity and that the DD genotype is more frequent and carries higher mortality in ARDS [2]. These observations underscore the involvement of ACE in ALI pathogenesis, although the nature of this involvement needs to be further clarified.

### Angiotensins

Angiotensins are a novel class of angiogenic growth factors whose contribution in sepsis and ALI pathophysiology has recently drawn increasing attention [38]. Angiotensin-1 is a Tie2 receptor agonist that reduces vascular permeability and inflammation, while angiotensin-2 has context-dependent effects acting as a Tie2 agonist or antagonist. Angiotensin-1 has been recently shown to exert an inhibitory effect on the synthesis and release of the proinflammatory and vasoconstrictor endothelin-1 by endothelial cells [39<sup>\*\*</sup>]. Angiotensin-2 acts mainly as a modulator of endothelial barrier disruption, promoting vascular leak in septic ALI [40] and hyperoxia-induced lung injury [41<sup>\*\*</sup>]; in contrast, an autocrine protective effect on activated endothelial cells that blocks vascular leak has also been suggested [42<sup>\*\*</sup>]. We have recently shown that serum angiotensin-2 levels in critically ill patients are increased in severe sepsis and correlate with serum TNF- $\alpha$  and disease severity. Human lung microvascular endothelial cells treated with LPS, TNF- $\alpha$ , and IL-6 responded by angiotensin-2 reduction, implying that human pulmonary endothelium may not be the source of increased angiotensin-2 in human sepsis. In contrast, LPS and TNF- $\alpha$  stimulated angiotensin-2 release by bovine lung microvascular endothelial cells, pointing to species-specific differences [38].

### Endothelium-related protective mechanisms and interventions in acute lung injury

Several compounds which are either synthesized by or act on endothelial cells have been used for lung injury treatment in experimental studies or in human trials. An overview of recent reports is given below.

#### Activated protein C

Activated protein C (APC) is used in the treatment of patients with severe sepsis. The mechanisms of action of this drug are under intense investigation. Endothelial protein C receptors (EPCRs) are expressed on endothelial, vascular smooth muscle, and immune cells. Receptor-deficient mice appeared more sensitive to LPS administration than receptor-expressing animals, while EPCRs offered protection when expressed on endothelial cells and not on hematopoietic cells, indicating that the mechanism of APC action is primarily endothelial cell related [43<sup>\*</sup>]. Animal studies by us and others, using various routes of APC administration and dosing regimens, documented lung-protective effects of the drug [44,45<sup>\*</sup>,46<sup>\*</sup>], despite apparent variations in the mode of APC action probably related to differences in experimental designs. Finally, of interest is the report of a homozygous 23 base pair insertion in the gene encoding EPCR in an 8-month-old boy with severe sepsis, pointing to the presence of a susceptibility-conferring mutation in this gene [47<sup>\*</sup>].

#### Antioxidants

In order to counteract excessive oxidant production, cells possess defensive antioxidant mechanisms. The transcription factor Nrf2 induces the expression of antioxidant enzymes. Nrf2 transcription can be activated by actin cytoskeletal remodeling in response to cyclic stretch, and by stimulation of the epidermal growth factor receptor signaling pathways [48]. A point mutation (-617C/A) in the promoter region of the Nrf2 gene, which decreased promoter activity, was identified in trauma patients, and was associated with increased risk for ARDS development [49<sup>\*</sup>]. The protective role of the antioxidant enzyme catalase was elucidated in an ischemia-reperfusion rat lung model; catalase conjugation to anti-ACE antibodies allowed lung endothelial cell targeting and attenuated lung injury [50<sup>\*\*</sup>].

#### Nitric oxide

A number of publications have dealt with the protective role of NO in lung injury. This is in contrast to data revealing permeability-increasing and cytotoxic effects. The discrepancy may be due to differences in experimental conditions, high versus low NO levels, as well as the site and source of NO release. Additionally, nitric oxide appears to exert either pro or antioxidative (i.e. harmful or protective) effects, depending on the type and concentration of ROS present in the microenvironment

[2] (see Figs 1 and 2). A most recent metaanalysis of studies on ARDS patients treated with inhaled NO revealed initial oxygenation improvement (a fact that supports the use of inhaled NO as a rescue therapy), no difference in clinical outcome, and an increased risk for renal dysfunction [31<sup>•</sup>]. The reason why the beneficial action of inhaled NO on the lungs appears neither sustained nor outcome improving is still puzzling and may be related, among other things, to dose–response changes with time and extrapulmonary organ toxicity. The observation that ALI patients with higher NO levels have reduced mortality, more ventilator-free days and more organ failure free days indicates that endogenous NO may, in fact, be cytoprotective due to immunomodulation or preservation of cell viability [51<sup>•</sup>]. Alternatively, since nitric oxide has bactericidal properties, patients with higher levels may be able to clear infections more efficiently.

The issue of the immune-modulating properties of endothelial nitric oxide synthase (eNOS)-derived NO was mechanistically addressed in a number of experimental studies using various lung injury models. eNOS-produced nitric oxide translated into: suppressed NF- $\kappa$ B activation in LPS models [52<sup>•</sup>]; lower activity levels of matrix metalloproteinase-9 (an enzyme which digests interstitial matrix) and thus a lower degree of fibrosis in mice receiving bleomycin [53<sup>•</sup>]; and abrogated PMN sequestration in mouse lungs primed with hypoxia and subsequently injured by ischemia–reperfusion [54].

#### Carbon monoxide

Low levels of carbon monoxide (CO) are released during enzymatic breakdown of heme by heme oxygenase-1 (HO-1) to perform important endothelial cell signaling functions. VEGF has been shown to induce HO-1 transcription and protect against hyperoxic ALI [55<sup>•</sup>]. Two additional studies revealed the protective effect of HO-1-produced CO and underscored the endothelial cell importance in hyperoxic injury [56<sup>•</sup>,57<sup>•</sup>]. In a model of murine thermal injury, CO donor drugs markedly reduced lung inflammation and edema, supporting the role of CO in lung endothelial cell cytoprotection and immune modulation against various insults [58<sup>•</sup>].

#### Adenosine

An important vasodilator, adenosine, has been shown to be released by mechanically stretched cells [59<sup>•</sup>]. Ectoapyrase (CD39) and ecto-5'-nucleotidase (CD73) are enzymes producing extracellular adenosine and were both found upregulated in VILI. Underexpression of both enzymes in mice resulted in worse VILI outcome, underscoring the role of adenosine in counteracting the deleterious effects of mechanical ventilation [59<sup>•</sup>]. The mode of adenosine protective action in ALI may be

related to preservation of intracellular energy pools [60] and prevention of apoptosis [61].

#### Cytoskeleton-stabilizing interventions

Newly developed pharmacologic agents have been reported to have endothelial barrier enhancing properties by strengthening endothelial cell–cell contacts via the actin cytoskeleton. This is presumably achieved by shifting the balance of actin polymerization towards cortical actin rings which reinforce cell–cell attachment, and away from stress fibers which pull cells away from each other [4]. Critical to these events is the function of small GTPases Rac and Cdc42. Compounds under study for this kind of effect are oxidized phospholipids, prostaglandins E<sub>2</sub> and I<sub>2</sub> and sphingosine-1 phosphate. These drugs have been tested in various lung injury models and were found to reduce lung edema and inflammation [62,63<sup>•</sup>,64,65].

#### Mechanisms of lung endothelial repair

Few studies addressed the vital issue of endothelial repair after the onset of injury. Since in clinical practice ALI is commonly diagnosed after its onset, understanding how to augment endothelial cell repair may be of critical importance to patient care. Zhao *et al.* [66<sup>•</sup>] showed that systemic LPS causes endothelial cell apoptosis in murine lungs. Transcription factor forkhead box M1 (FoxM1), known for its role in DNA replication and mitosis, is crucial to endothelial regeneration after LPS [66<sup>•</sup>]. In a clinically relevant animal experiment, recombinant VEGF administration enhanced alveolar and vascular regeneration in hyperoxic rat pups, albeit at the expense of transiently increasing permeability [67<sup>•</sup>]. Additionally, Quadri and Bhattacharya [68] used hydrogen peroxide to induce loss of cell–cell contacts in cultured endothelial cell monolayers, and showed that endothelial restoration was dependent on activation of focal adhesion kinase, an enzyme found in focal adhesions.

#### Endothelial–epithelial interactions

Kinniry *et al.* [69<sup>•</sup>] described a novel role for KL(4)-surfactant in ALI protection. KL(4)-surfactant, containing the novel peptide sinapultide, administered intranasally diminished PMN infiltration in lungs of mice challenged with hyperoxia or LPS. This effect appeared related to blockade of PMN transmigration across the endothelium.

#### Novel lung-targeting therapies

Two already presented studies demonstrated the feasibility of lung targeting of genes or proteins with endothelial cell protective properties. In the work by McCarter *et al.* [39<sup>••</sup>], rat dermal fibroblasts transfected *in vitro* with the gene coding for angiopoietin-1 were injected intravenously to the intact animal and were rapidly incorporated in the lung microvessel walls. ALI was induced 24 h later by LPS intratracheal instillation. Animals overexpressing angiopoietin-1 sustained less lung injury than controls, a

fact attributed to the barrier-sealing effect of angiopoietin-1 and/or to endothelin-1 decreased expression. In the second study, the antioxidant enzyme catalase was conjugated to monoclonal antibodies against rat ACE, thus concentrating the enzyme on lung endothelial cells. Rats treated with this conjugate exhibited a significant attenuation of ischemia–reperfusion lung injury [50\*\*].

## Conclusion

The endothelial cell is central to the pathogenesis of ALI. Endothelial-targeted interventions are able to prevent lung injury in several experimental models. Keeping in mind that there is usually a long way to go from bench to bedside, some of these modalities may in the long term prove to be useful in the clinical setting and improve the management and prognosis of this devastating complication.

## Acknowledgements

This work was supported by the Thorax Foundation and a grant from the Hellenic Society of Critical Care.

## References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- of outstanding interest

Additional references related to this topic can also be found in the Current World Literature section in this issue (pp. 104–105).

- 1 Aird WC. Phenotypic heterogeneity of the endothelium: II. Representative vascular beds. *Circ Res* 2007; 100:174–190. Comprehensively illustrated detailed review on endothelium heterogeneity.
- 2 Orfanos SE, Mavrommati I, Korovesi I, Roussos C. Pulmonary endothelium in acute lung injury: from basic science to the critically ill. *Intensive Care Med* 2004; 30:1702–1714.
- 3 Mehta D, Malik AB. Signaling mechanisms regulating endothelial permeability. *Physiol Rev* 2006; 86:279–367.
- 4 Dudek SM, Garcia JG. Cytoskeletal regulation of pulmonary vascular permeability. *J Appl Physiol* 2001; 91:1487–1500.
- 5 Reutershan J, Stockton R, Zarbock A, *et al.* Blocking p21-activated kinase reduces lipopolysaccharide-induced acute lung injury by preventing polymorphonuclear leukocyte infiltration. *Am J Respir Crit Care Med* 2007; 175:1027–1035.
- 6 Rossi JL, Velentza AV, Steinhorn DM, *et al.* MLCK210 gene knockout or kinase inhibition preserves lung function following endotoxin-induced lung injury in mice. *Am J Physiol Lung Cell Mol Physiol* 2007; 292:L1327–L1334.
- 7 Ralay Ranaivo H, Carusio N, Wangenstein R, *et al.* Protection against endotoxic shock as a consequence of reduced nitrosative stress in MLCK210-null mice. *Am J Pathol* 2007; 170:439–446. This work provides evidence for a link between endothelial cell cytoskeletal remodeling and regulation of proinflammatory gene expression in ALL.
- 8 Bogatcheva NV, Adyshev D, Mambetsariev B, *et al.* Involvement of microtubules, p38, and Rho kinases pathway in 2-methoxyestradiol-induced lung vascular barrier dysfunction. *Am J Physiol Lung Cell Mol Physiol* 2007; 292:L487–L499.
- 9 Mirzapioazova T, Kolosova IA, Moreno L, *et al.* Suppression of endotoxin-induced inflammation by taxol. *Eur Respir J* 2007; 30:429–435.
- 10 Su G, Hodnett M, Wu N, *et al.* Integrin alphavbeta5 regulates lung vascular permeability and pulmonary endothelial barrier function. *Am J Respir Cell Mol Biol* 2007; 36:377–386.
- 11 Lu Q, Harrington EO, Jackson H, *et al.* Transforming growth factor-beta1-induced endothelial barrier dysfunction involves Smad2-dependent p38 activation and subsequent RhoA activation. *J Appl Physiol* 2006; 101: 375–384.

- 12 Georgieva GS, Kurata S, Ikeda S, *et al.* Nonischemic lung injury by mediators from unilateral ischemic reperfusion lung: ameliorating effect of tumor necrosis factor-alpha-converting enzyme inhibitor. *Shock* 2007; 27:84–90.

This work demonstrates remote organ effects of TNF- $\alpha$  secreted from lungs undergoing ischemia/reperfusion.

- 13 Angelini DJ, Hyun SW, Grigoryev DN, *et al.* TNF-alpha increases tyrosine phosphorylation of vascular endothelial cadherin and opens the paracellular pathway through fyn activation in human lung endothelia. *Am J Physiol Lung Cell Mol Physiol* 2006; 291:L1232–L1245.

Novel mechanism of endothelial cell barrier disruption by TNF- $\alpha$ .

- 14 Alvarez DF, King JA, Weber D, *et al.* Transient receptor potential vanilloid 4-mediated disruption of the alveolar septal barrier: a novel mechanism of acute lung injury. *Circ Res* 2006; 99:988–995.

This work clarifies a mode of Ca<sup>2+</sup> influx from the extracellular space in endothelial cells, which induces permeability increases in ALL.

- 15 Parthasarathi K, Ichimura H, Monma E, *et al.* Connexin 43 mediates spread of Ca<sup>2+</sup>-dependent proinflammatory responses in lung capillaries. *J Clin Invest* 2006; 116:2193–2200.

This article describes Ca<sup>2+</sup>-mediated spread of inflammatory signals along adjacent endothelial cells interconnected via gap junctions, thus propagating inflammation in ALL.

- 16 Chatterjee A, Dimitropoulou C, Drakopanayiotakis F, *et al.* Heat shock protein 90 inhibitors prolong survival, attenuate inflammation and reduce lung injury in murine sepsis. *Am J Respir Crit Care Med* 2007; 176:667–675.

Work showing the protective effect of HSP-90 inhibitors in sepsis/ALI, bearing potential clinical implications.

- 17 Kim KS, Rajagopal V, Gonsalves C, *et al.* A novel role of hypoxia-inducible factor in cobalt chloride- and hypoxia-mediated expression of IL-8 chemokine in human endothelial cells. *J Immunol* 2006; 177:7211–7224.

- 18 Yoshida K, Kondo R, Wang Q, Doerschuk CM. Neutrophil cytoskeletal rearrangements during capillary sequestration in bacterial pneumonia in rats. *Am J Respir Crit Care Med* 2006; 174:689–698.

- 19 Perkowski S, Scherpereel A, Murciano JC, *et al.* Dissociation between alveolar transmigration of neutrophils and lung injury in hyperoxia. *Am J Physiol Lung Cell Mol Physiol* 2006; 291:L1050–L1058.

- 20 Gao XP, Zhu X, Fu J, *et al.* Blockade of class IA phosphoinositide 3-kinase in neutrophils prevents NADPH oxidase activation- and adhesion-dependent inflammation. *J Biol Chem* 2007; 282:6116–6125.

This work characterizes a molecular pathway of ROS production by TNF- $\alpha$ -stimulated PMNs with implications on endothelial permeability.

- 21 Kiefmann R, Heckel K, Schenkat S, *et al.* Role of p-selectin in platelet sequestration in pulmonary capillaries during endotoxemia. *J Vasc Res* 2006; 43:473–481.

- 22 Zarbock A, Singbartl K, Ley K. Complete reversal of acid-induced acute lung injury by blocking of platelet–neutrophil aggregation. *J Clin Invest* 2006; 116:3211–3219.

TxA<sub>2</sub> secreted by complexes of platelet–PMN is critical in inducing vascular permeability and inflammation.

- 23 Khan SY, Kelher MR, Heal JM, *et al.* Soluble CD40 ligand accumulates in stored blood components, primes neutrophils through CD40, and is a potential cofactor in the development of transfusion-related acute lung injury. *Blood* 2006; 108:2455–2462.

Soluble CD40 ligand-activated PMNs induce endothelial alterations in transfusion-related acute lung injury.

- 24 Pero RS, Borchers MT, Spicher K, *et al.* G $\alpha$ phai2-mediated signaling events in the endothelium are involved in controlling leukocyte extravasation. *Proc Natl Acad Sci U S A* 2007; 104:4371–4376.

Insight in endothelial cell signaling pathways that regulate PMN transmigration.

- 25 Orrington-Myers J, Gao X, Kouklis P, *et al.* Regulation of lung neutrophil recruitment by VE-cadherin. *Am J Physiol Lung Cell Mol Physiol* 2006; 291:L764–L771.

Vascular endothelial cadherin signaling regulates NF- $\kappa$ B activation and adhesion molecule expression.

- 26 Abdounour RE, Peng X, Finigan JH, *et al.* Mechanical stress activates xanthine oxidoreductase through MAP kinase-dependent pathways. *Am J Physiol Lung Cell Mol Physiol* 2006; 291:L345–L353.

- 27 Flaishon R, Szold O, Weinbroum AA. Acute lung injury following pancreas ischaemia-reperfusion: role of xanthine oxidase. *Eur J Clin Invest* 2006; 36:831–837.

- 28 Usatyuk PV, Parinandi NL, Natarajan V. Redox regulation of 4-hydroxy-2-nonenal-mediated endothelial barrier dysfunction by focal adhesion, adherens, and tight junction proteins. *J Biol Chem* 2006; 281:35554–35566.

- 29** Silliman CC, Curtis BR, Kopko PM, *et al.* Donor antibodies to HNA-3a • implicated in TRALI reactions prime neutrophils and cause PMN-mediated damage to human pulmonary microvascular endothelial cells in a two-event in vitro model. *Blood* 2007; 109:1752–1755.
- Transfusion-related acute lung injury requires a 'two-hit' model of preexisting endothelial cell stimulation followed by oxidant-mediated endothelial cell toxicity from PMNs activated by antibodies in donor blood.
- 30** Gorbunov NV, Das DK, Goswami SK, *et al.* Spatial coordination of cell-adhesion molecules and redox cycling of iron in the microvascular inflammatory response to pulmonary injury. *Antioxid Redox Signal* 2007; 9:483–495.
- 31** Adhikari NK, Burns KE, Friedrich JO, *et al.* Effect of nitric oxide on oxygenation and mortality in acute lung injury: systematic review and meta-analysis. *BMJ* 2007; 334:757–758.
- Most recent metaanalysis arguing against the routine use of inhaled nitric oxide in ALI/ARDS due to lack of clinical outcome improvement and potential renal toxicity.
- 32** Ader F, Le Berre R, Lancel S, *et al.* Inhaled nitric oxide increases endothelial permeability in *Pseudomonas aeruginosa* pneumonia. *Intensive Care Med* 2007; 33:503–510.
- 33** Shelton JL, Wang L, Cepinskas G, *et al.* Inducible NO synthase (iNOS) in human neutrophils but not pulmonary microvascular endothelial cells (PMVEC) mediates septic protein leak in vitro. *Microvasc Res* 2007; 74:23–31.
- 34** Shields CJ, Delaney CP, Winter DC, *et al.* Induction of nitric oxide synthase is a key determinant of progression to pulmonary injury in experimental pancreatitis. *Surg Infect (Larchmt)* 2006; 7:501–511.
- 35** He X, Han B, Mura M, *et al.* Angiotensin-converting enzyme inhibitor captopril prevents oleic acid-induced severe acute lung injury in rats. *Shock* 2007; 28:106–111.
- 36** Jerng JS, Hsu YC, Wu HD, *et al.* Role of the renin–angiotensin system in ventilator-induced lung injury: an in vivo study in a rat model. *Thorax* 2007; 62:527–535.
- This article demonstrates increased levels of angiotensin II in a rat VILI model and lung protection by ACE inhibition.
- 37** Jerng JS, Yu CJ, Wang HC, *et al.* Polymorphism of the angiotensin-converting enzyme gene affects the outcome of acute respiratory distress syndrome. *Crit Care Med* 2006; 34:1001–1006.
- This work shows that ACE D allele carriers have increased ARDS mortality.
- 38** Orfanos SE, Kotanidou A, Glynos C, *et al.* Angiotensin-2 is increased in severe sepsis: correlation with inflammatory mediators. *Crit Care Med* 2007; 35:199–206.
- 39** McCarter SD, Mei SH, Lai PF, *et al.* Cell-based angiotensin-1 gene therapy •• for acute lung injury. *Am J Respir Crit Care Med* 2007; 175:1014–1026.
- Skin fibroblasts genetically engineered to secrete high levels of angiotensin-1 can be incorporated in lung vessel walls and confer protection against ALI.
- 40** Parikh SM, Mammoto T, Schultz A, *et al.* Excess circulating angiotensin-2 may contribute to pulmonary vascular leak in sepsis in humans. *PLoS Med* 2006; 3:e46:0356–0370.
- 41** Bhandari V, Choo-Wing R, Lee CG, *et al.* Hyperoxia causes angiotensin-•• 2-mediated acute lung injury and necrotic cell death. *Nat Med* 2006; 12:1287–1293.
- Toxic effects of hyperoxia on lung tissue are mediated by angiotensin-2 secretion.
- 42** Daly C, Pasnikowski E, Burova E, *et al.* Angiotensin-2 functions as an •• autocrine protective factor in stressed endothelial cells. *Proc Natl Acad Sci U S A* 2006; 103:15491–15496.
- Evidence for the novel protective properties of angiotensin-2.
- 43** Zheng X, Li W, Song Y, *et al.* Nonhematopoietic EPCR regulates • the coagulation and inflammatory responses during endotoxemia. *J Thromb Haemost* 2007; 5:1394–1400.
- Endothelial protein C receptors on endothelial cells and not hematopoietic cells protect mice from sepsis-induced mortality and ALI.
- 44** Maybauer MO, Maybauer DM, Fraser JF, *et al.* Recombinant human activated protein C improves pulmonary function in ovine acute lung injury resulting from smoke inhalation and sepsis. *Crit Care Med* 2006; 34:2432–2438.
- 45** Kotanidou A, Loutrari H, Papadomichelakis E, *et al.* Inhaled activated protein C • attenuates lung injury induced by aerosolized endotoxin in mice. *Vascul Pharmacol* 2006; 45:134–140.
- This work shows the protective effect of APC topical administration in a model of direct ALI, by influencing endothelial cell–PMN interactions.
- 46** Slofstra SH, Groot AP, Maris NA, *et al.* Inhalation of activated protein C • inhibits endotoxin-induced pulmonary inflammation in mice independent of neutrophil recruitment. *Br J Pharmacol* 2006; 149:740–746.
- This work offers evidence on the protective effect of topical administration of APC in a model of direct ALI via PMN-independent pathway(s).
- 47** Kendirli T, Ciftci E, Ince E, *et al.* Homozygous 23-bp insertion of endothelial • protein C receptor gene in a child with fatal sepsis. *Pediatr Hematol Oncol* 2007; 24:199–204.
- A case reporting a mutation of the endothelial protein C receptor possibly associated with poor outcome.
- 48** Papaiahgari S, Yerrapureddy A, Hassoun PM, *et al.* EGFR-activated signaling and actin remodeling regulate cyclic stretch-induced NRF2-ARE activation. *Am J Respir Cell Mol Biol* 2007; 36:304–312.
- 49** Marzec JM, Christie JD, Reddy SP, *et al.* Functional polymorphisms in the • transcription factor NRF2 in humans increase the risk of acute lung injury. *FASEB J* 2007; 21:2237–2246.
- Genetic polymorphism in the NRF2 promoter increases the risk of ALI in trauma patients.
- 50** Nowak K, Weih S, Metzger R, *et al.* Immunotargeting of catalase to lung •• endothelium via antiangiotensin-converting enzyme antibodies attenuates ischemia–reperfusion injury of the lung in vivo. *Am J Physiol Lung Cell Mol Physiol* 2007; 293:L162–L169.
- This work describes a novel approach to concentrate catalase on lung endothelial cells by conjugating it to antibodies against ACE.
- 51** McClintock DE, Ware LB, Eisner MD, *et al.* Higher urine nitric oxide is • associated with improved outcomes in patients with acute lung injury. *Am J Respir Crit Care Med* 2007; 175:256–262.
- Unexpected finding that ALI patients with higher nitric oxide levels have better outcomes.
- 52** Garrean S, Gao XP, Brovkovich V, *et al.* Caveolin-1 regulates NF-kappaB • activation and lung inflammatory response to sepsis induced by lipopolysaccharide. *J Immunol* 2006; 177:4853–4860.
- Excess eNOS-derived nitric oxide is protective in septic ALI in mice.
- 53** Yoshimura S, Nishimura Y, Nishiuma T, *et al.* Overexpression of nitric oxide • synthase by the endothelium attenuates bleomycin-induced lung fibrosis and impairs MMP-9/TIMP-1 balance. *Respirology* 2006; 11:546–556.
- This work shows that transgenic endothelial cell-specific eNOS expression counteracts the effects of bleomycin.
- 54** Kaminski A, Kasch C, Zhang L, *et al.* Endothelial nitric oxide synthase mediates protective effects of hypoxic preconditioning in lungs. *Respir Physiol Neurobiol* 2007; 155:280–285.
- 55** Siner JM, Jiang G, Cohen ZI, *et al.* VEGF-induced heme oxygenase-1 confers • cytoprotection from lethal hyperoxia in vivo. *FASEB J* 2007; 21:1422–1432.
- Refs [55\*–57\*] offer information on the endothelial cell protective actions of carbon monoxide in hyperoxia.
- 56** Zhang X, Shan P, Jiang G, *et al.* Endothelial STAT3 is essential for the • protective effects of HO-1 in oxidant-receptor lung injury. *FASEB J* 2006; 20:2156–2158.
- Refs [55\*–57\*] offer information on the endothelial cell protective actions of carbon monoxide in hyperoxia.
- 57** Wang X, Wang Y, Kim HP, *et al.* Carbon monoxide protects against hyperoxia- • induced endothelial cell apoptosis by inhibiting reactive oxygen species formation. *J Biol Chem* 2007; 282:1718–1726.
- Refs [55\*–57\*] offer information on the endothelial cell protective actions of carbon monoxide in hyperoxia.
- 58** Sun B, Sun H, Liu C, *et al.* Role of CO-releasing molecules liberated CO in • attenuating leukocytes sequestration and inflammatory responses in the lung of thermally injured mice. *J Surg Res* 2007; 139:128–135.
- This work provides in-vivo evidence for the beneficial effect of carbon monoxide donors in ALI induced by burns.
- 59** Eckle T, Füllbier L, Wehrmann M, *et al.* Identification of ectonucleotidases • CD39 and CD73 in innate protection during acute lung injury. *J Immunol* 2007; 178:8127–8137.
- Adenosine produced by enzymes CD39 and CD73 protects mice from ALI.
- 60** Heller AR, Rothermel J, Weigand MA, *et al.* Adenosine A1 and A2 receptor agonists reduce endotoxin-induced cellular energy depletion and oedema formation in the lung. *Eur J Anaesthesiol* 2007; 24:258–266.
- 61** Rivo J, Zeira E, Galun E, *et al.* Attenuation of reperfusion lung injury and apoptosis by A2A adenosine receptor activation is associated with modulation of Bcl-2 and Bax expression and activation of extracellular signal-regulated kinases. *Shock* 2007; 27:266–273.
- 62** Birukova AA, Zagranichnaya T, Fu P, Alekseeva E, *et al.* Prostaglandins PGE(2) and PGI(2) promote endothelial barrier enhancement via PKA and Epac1/Rap1-dependent Rac activation. *Exp Cell Res* 2007; 313:2504–2520.
- 63** Birukova AA, Malyukova I, Poroyko V, Birukov KG. Paxillin-beta–catenin • interactions are involved in Rac/Cdc42-mediated endothelial barrier-protective response to oxidized phospholipids. *Am J Physiol Lung Cell Mol Physiol* 2007; 293:L199–L211.
- Oxidized phospholipids are compounds which promote sealing of endothelial junctions and enhance the endothelial barrier tightness.

- 64** Birukova AA, Fu P, Chatchavalvanich S, *et al.* Polar head groups are important for barrier-protective effects of oxidized phospholipids on pulmonary endothelium. *Am J Physiol Lung Cell Mol Physiol* 2007; 292:L924–L935.
- 65** Okazaki M, Kreisel F, Richardson SB, *et al.* Sphingosine 1-phosphate inhibits ischemia reperfusion injury following experimental lung transplantation. *Am J Transplant* 2007; 7:751–758.
- 66** Zhao YY, Gao XP, Zhao YD, *et al.* Endothelial cell-restricted disruption of FoxM1 impairs endothelial repair following LPS-induced vascular injury. *J Clin Invest* 2006; 116:2333–2343.
- Sepsis induces lung endothelial cell apoptosis; subsequent proliferation of surviving endothelial cells, dependent on transcription factor FoxM1, is essential to recovery from ALL.
- 67** Kunig AM, Balasubramaniam V, Markham NE, *et al.* Recombinant human VEGF treatment transiently increases lung edema but enhances lung structure after neonatal hyperoxia. *Am J Physiol Lung Cell Mol Physiol* 2006; 291:L1068–L1078.
- Exogenous VEGF increases permeability but restores vascular development in neonatal hyperoxia.
- 68** Quadri SK, Bhattacharya J. Resealing of endothelial junctions by focal adhesion kinase. *Am J Physiol Lung Cell Mol Physiol* 2007; 292:L334–L342.
- 69** Kinniry P, Pick J, Stephens S, *et al.* KL4-surfactant prevents hyperoxic and LPS-induced lung injury in mice. *Pediatr Pulmonol* 2006; 41:916–928.
- Surfactant component KL(4) from epithelial cells protects endothelial cells in endotoxemic mice.